J.J. College of Arts and Science (Autonomous)

Sivapuram, Pudukkottai - 622 422

PG and Research Department of Microbiology

M.Sc., Microbiology

Syllabus based on

TANSCHE Regulations on Learning Outcomes - Based Curriculum Frame Work for Post Graduate Education

[Those who Join 2023 - 2024 Onwards]

J.J. College of Arts and Science (Autonomous)
Sivapuram Post, Pudukkottai

Department of Microbiology

Programme Outcomes – PG (2023-2024)

PO1: Problem Solving Skill: Apply knowledge of Management theories and Human Resource practices to solve business problems through research in Global context.

PO2: Decision Making Skill: Foster analytical and critical thinking abilities for data-based decision-making.

PO3: Ethical Value: Ability to incorporate quality, ethical and legal value-based perspectives to all organizational activities.

PO4: Communication Skill: Ability to develop communication, managerial and interpersonal skills.

PO5: Individual and Team Leadership Skill: Capability to lead themselves and the team to achieve organizational goals.

PO6: Employability Skill: Inculcate contemporary business practices to enhance employability skills in the competitive environment.

PO7: Entrepreneurial Skill: Equip with skills and competencies to become an entrepreneur.

PO8: Contribution to Society: Succeed in career endeavors and contribute significantly to society.

PO9: Multicultural competence: Possess knowledge of the values and beliefs of multiple cultures and a global perspective.

PO10: Moral and ethical awareness/reasoning: Ability to embrace moral/ethical values in conducting one's life.

J.J. College of Arts and Science (Autonomous)

Sivapuram Post, Pudukkottai

Department of Microbiology

Programme Specific Outcomes – PG (2023-2024)

PSO1: Placement: To prepare the students who will demonstrate respectful engagement with others' ideas, behaviours, beliefs and apply diverse frames of reference to decisions and actions.

PSO 2: Entrepreneur: To create effective entrepreneurs by enhancing their critical thinking, problem solving, decision making and leadership skill that will facilitate start-ups and high potential organizations.

PSO3: Research and Development: Design and implement HR systems and practices grounded in research that comply with employment laws, leading the organization towards growth and development.

PSO4: Contribution to Business World: To produce employable, ethical and innovative professionals to sustain in the dynamic business world.

PSO5: Contribution to the Society: To contribute to the development of the society by collaborating with stakeholders for mutual benefit.

J.J. College of Arts and Science (Autonomous)

Department of Microbiology

Proposed Course Structure based on TANSCHE and UGC - LOCF
(Choice Based Credit System)
(Applicable for the Candidates admitted from academic year 2023-2024 Onwards)

M.Sc. Microbiology - Semester I

Sl.No.	Course Code	Course	Overall Credits	Total Contact Hours/Week	Marks		
					CIA	ESE	Total
Semester I							
1	P1R3MBCC1	General Microbiology & Microbial Diversity	5	7	25	75	100
2	P1R3MBCC2	Immunology, Immunomics & Microbial Genetics	5	7	25	75	100
3	P1R3MBCC3P	Practical - I Immunology, Immunomics & Microbial Genetics	4	6	40	60	100
4	P1R3MBDSE 1:1 / 1:2 / 1:3	Anyone from the List	3	5	25	75	100
5	P1R3MBDSE 2:1 / 2:2 / 2:3	Anyone from the List	3	5	25	75	100
		Total	**20**	**30**			**500**

SEMESTER I

GENERAL MICROBIOLOGY AND MICROBIAL DIVERSITY

Course Code: P1R3MBCC1	**Credit:** 5
Category: Core Course	**Hrs/Week:** 7, **Total Inst.Hrs:** 105
Nature of the Course: Skill Development	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Acquire knowledge on the principles of different types of microscopes and their applications.
2. Compare and contrast the structure of bacteria and fungi. Illustrate nutritional requirements and growth in bacteria.
3. Exemplify, isolate and cultivate microalgae from diverse environmental sources.
4. Explain various pure culture techniques and discuss sterilization methods.
5. Discuss the importance and conservation of microbial diversity.

Unit I: **21 hrs**

History and Scope of Microbiology. Microscopy – Principles and applications. Types of Microscopes - Transmission electron microscope (TEM) and Scanning electron microscope (SEM). Sample preparation for SEM & TEM. Atomic force, Confocal microscope. Micrometry – Stage, Ocular and its applications.

Unit II: **21 hrs**

Bacterial Structure, properties and biosynthesis of cellular components – Cell wall. Actinomycetes and Fungi - Distribution, morphology, classification, reproduction and economic importance. Sporulation. Growth and nutrition - Nutritional requirements, Growth curve, Kinetics of growth, Batch culture, Synchronous growth, Measurement of growth and factors affecting growth.

Unit III: **21 hrs**

Algae - Distribution, morphology, classification, reproduction and economic importance. Isolation of algae from soil and water. Media and methods used for culturing algae, Strain selection and large-scale cultivation. Life cycle - *Chlamydomonas*, *Volvox Spirogyra* (Green

algae), *Nostoc* (Cyanobacteria) *Ectocarpus*, *Sargassum* (Brown algae), *Polysiphonia*, *Batrachospermum* (Red algae).

Unit IV: **21 hrs**

Microbial techniques - Safety guidelines in Microbiology Laboratories. Sterilization, Disinfection and its validation. Staining methods – Simple, Differential and Special staining. Automated Microbial identification systems - Pure cultures techniques – Cultivation of Anaerobic organisms. Maintenance and preservation of pure cultures. Culture collection centres - National and International.

Unit V: **21 hrs**

Biodiversity - Introduction to microbial biodiversity – Thermophiles - Classification, Thermophilic Archaebacteria and its applications. Methanogens - Classification, Habitats, applications. Alkaliphiles and Acidophiles - Classification, discovery basin, its cell wall and membrane. Barophiles - Classification and its applications. Halophiles - Classification, discovery basin, cell walls and membranes – purple membrane, compatible solutes, Osmoadaptation / halotolerance - Applications of halophiles. Conservation of Biodiversity.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Examine various microbes employing the microscopic techniques learnt. Measure and compare the size of microbes.	PO1, PO4, PO11
CO2	Differentiate and appreciate the anatomy of various microbes. Plan the growth of microbes for different environmental conditions.	PO1, PO4
CO3	Identify and cultivate the algae understanding their habitat. Analyze the morphology, classify and propagate depending on its economic importance.	PO7, PO8, PO9
CO4	Create aseptic conditions by following good laboratory practices.	PO3, PO4,PO7
CO5	Categorize and cultivate a variety of extremophiles following standard protocols for industrial applications.	PO5, PO7, PO8, PO9

Text Books

1. Kanunga R. (2017). Ananthanarayanan and Panicker's Text book of Microbiology. (10th Edition). Universities Press (India) Pvt. Ltd.

2. Chan E.C.S., Pelczar M. J. Jr. and Krieg N. R. (2010). Microbiology. (5th Edition). McGraw Hill. Inc, New York.
3. Prescott L. M., Harley J. P. and Klein D. A. (2004). Microbiology. (6th Edition). McGraw - Hill company, New York.
4. White D. Drummond J. and Fuqua C. (2011). The Physiology and Biochemistry of Prokaryotes, Oxford University Press, Oxford, New York.
5. Dubey R.C. and Maheshwari D. K. (2009). Textbook of Microbiology. S. Chand, Limited.

Reference Books

1. Tortora G. J., Funke B. R. and Case C. L. (2015). Microbiology: An Introduction (12th Edition).Pearson, London, United Kingdom
2. Webster J. and Weber R.W.S. (2007). Introduction to Fungi. (3rd Edition). Cambridge University Press, Cambridge.
3. Schaechter M. and Leaderberg J. (2004). The Desk encyclopedia of Microbiology. Elseiver Academic Press, California.
4. Ingraham, J.L. and Ingraham, C.A. (2000) Introduction to Microbiology. (2nd Edition). Books / Cole Thomson Learning, UK.
5. Madigan M. T., Bender K.S., Buckley D. H. Sattley W. M. and Stahl (2018) Brock Biology of Microorganisms. (15th Edition). Pearson.

Web Resources

1. http://sciencenetlinks.com/tools/microbeworld
2. https://www.microbes.info/
3. https://www.asmscience.org/VisualLibrary
4. https://open.umn.edu/opentextbooks/BookDetail.aspx?bookId=404
5. https://www.grsmu.by/files/file/university/cafedry//files/essential_microbiology.pdf

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO 1	M			M							S			
CO 2	L			S										
CO 3							S	S	M					
CO 4			S	S			S							
CO 5					S		S	S	S					

IMMUNOLOGY, IMMUNOMICS AND MICROBIAL GENETICS

Course Code: P1R3MBCC2	**Credit:** 5
Category: Core Course	**Hrs/Week: 7**, **Total Inst.Hrs:** 105
Nature of the Course: Skill Development	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Discuss immunity, organs and cells involved in immunity. Compare the types of antigens and their properties.
2. Describe immunoglobulin and its types. Categorize MHC and understand its significance.
3. Elucidate the mechanisms of different hypersensitivity reactions. List out the Vaccines and discuss their development.
4. Acquire knowledge the structure DNA in prokaryotes and eukaryotes.
5. Explain out gene transfer studies in microbes.

Unit I: **21 hrs**

Introduction to biology of the immune system – Cells and organs of Immune System. T and B lymphocytes – Origin, development, differentiation, lymphocyte subpopulation in humans. Innate immunity- Complement, Acquired immunity – Active and Passive immunity. Antigens - features associated with antigenicity and immunogenicity. Basis of antigen specificity. MHC genes and products, Structure of MHC molecules, Genetics of HLA Systems – Antigens and HLA typing.

Unit II: **21 hrs**

Immunoglobulins. Theories of antibody production. Monoclonal and polyclonal antibodies. Complement system – mode of activation- Classical, Alternate and Lectin pathways, biological functions. Antigen recognition – TCR, Diversity of TCR, T cell surface alloantigens, lymphocyte activation, clonal proliferation and differentiation. Physiology of acquired immune response – various phases of HI, CMI – Cell mediated cytotoxicity, DTH response.

Unit III: **21 hrs**

Hypersensitivity – Types and mechanisms, Autoimmunity, Tumor Immunity and Transplantation immunology. Immunodeficiency-Primary immunodeficiency and Secondary immunodeficiencies. Genetics of Immunohematology – Genetic basis and significance of ABO and other minor blood groups in humans, Bombay blood group, Secretors and Non-secretors, Rh System and genetic basis of D- antigens. Agglutination - Hemagglutination - Hemagglutination inhibition. Labeled Assay- Immunofluorescence assay, Radio immunoassay, FISH, ELISA. Flow cytometry. Immune regulation mechanisms – immuno-induction, immuno- suppression, immuno-tolerance, immuno-potentiation, Immunomodulation. Role of cytokines, lymphokines and chemokines. Immunomics - Introduction and Applications. Antigen engineering for better immunogenicity and use for vaccine development-multiepitope vaccines. Reverse vaccinology.

Unit IV: **21 hrs**

Structural of prokaryotic and eukaryotic genome. Introduction to prokaryotic genomic structure, Eukaryotic Genome - Structure of chromatin, chromosome, centromere, telomere, nucleosome. Modifications- methylation, acetylation, phosphorylation and its effect on structure and function of chromatin, DNA methylation and gene imprinting, organelle genome.

Unit V: **21 hrs**

Gene Transfer Mechanisms- Conjugation and its uses. Transduction, Generalized and Specialized, Transformation– Natural Competence and Transformation. Transposition and Types of Transposition reactions. Insertion sequences, complex and compound transposons – T10, T5, and Retroposon. Mechanism – Transposons of *E. coli*, Bacteriophage and Yeast. Importance of transposable elements in horizontal transfer of genes and evolution.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Categorize the immune response to a variety of antigens. Identify different immune cells involved in immunity.	PO1, PO4, PO6, PO7, PO9
CO2	Justify the significance of MHC molecules in immune response and antibody production.	PO1, PO4, PO5,PO6, PO9
CO3	Design antibodies and evaluate immunological assays in patient samples.	PO4, PO6, PO7, PO8, PO9, PO10
CO4	Analyze genomic DNA of prokaryotes and eukaryotes.	PO4,PO5, PO6, PO7, PO9, PO10
CO5	Summarize gene transfer mechanisms for experimental study.	PO4,PO5, PO6, PO7, PO9, PO10

Text Books

1. Coico R., Sunshine G. and Benjamini E. (2003). Immunology – A Short Course. (5th Edition). Wiley-Blackwell, New York.
2. Owen J. A., Punt J., Stranford S. A. and Kuby J. (2013). Immunology, (7th Edition). W. H. Freeman and Company, New York.
3. Abbas A. K., Lichtman A. H. and Pillai S. (2021). Cellular and Molecular Immunology. (10th Edition). Elsevier.
4. Malacinski G.M. (2008). Freifelder's Essentials of Molecular Biology. (4th Edition). Narosa Publishing House, New Delhi.
5. Gardner E. J. Simmons M. J. and Snusted D.P. (2006). Principles of Genetics. (8th Edition). Wiley India Pvt. Ltd.

Reference Books

1. Travers J. (1997). Immunobiology - The Immune System in Health and Disease. (3rd Edition). Current Biology Ltd. New York.

2. Delves P.J., Martin S., Burton D. R. and Roitt I. M. (2006). Roitt's Essential Immunology. (11th Edition). Wiley-Blackwell.

3. Hay F. C. and Westwood O. M. R. (2002). Practical Immunology (4th Edition). Wiley-Blackwell.

4. Glick B. R. and Patten C.L. (2018). Molecular Biotechnology – Principles and Applications of Recombinant DNA. (5th Edition). ASM Press.

5. Russell P.J. (2010). Genetics - A Molecular Approach. (3rd Edition). Pearson New International Edition.

Web Resources

1. https://www.ncbi.nlm.nih.gov/books/NBK279395/
2. https://med.stanford.edu/immunol/phd-program/ebook.html
3. https://ocw.mit.edu/courses/hst-176-cellular-and-molecular-immunology-fall-2005/pages/lecture-notes/
4. [PDF] Lehninger Principles of Biochemistry (8th Edition) By David L. Nelson and Michael M. Cox Book Free Download - StudyMaterialz.in
5. https://microbenotes.com/gene-cloning-requirements-principle-steps-applications/

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S			M		M	S		S					
CO2	S			S	M	S			S					
CO3				S		S	S	S	S	M				
CO4				S	M	S	M		S	M				
CO5				S	M	S	M		S	S				

PRACTICAL I

GENERAL MICROBIOLOGY AND MICROBIAL DIVERSITY & IMMUNOLOGY, IMMUNOMICS AND MICROBIAL GENETICS

Course Code: P1R3MBCC3P	**Credit:** 4
Category: Core Course – Practical I	**Hrs/Week:** 6
Nature of the Course: Skill Development	**Marks:** CIA: 40+ EXT: 60 = 100

Course Objectives

1. Gain knowledge on the fundamentals, handling and applications of microscopy, sterilization methods. Identify microbes by different staining methods.
2. Prepare media for bacterial growth. Discuss plating and growth measurement techniques.
3. Acquire adequate skills to perform blood grouping and serological reactions.
4. Provide fundamental skills in preparation, separation and purification of immunoglobulin.
5. Apply the knowledge of molecular biology skills in clinical diagnosis.

1. Microscopic Techniques: Light microscopy: Hay infusion broth. Wet mount to show different types of microbes, hanging drop.
2. Dark field microscopy – Motility of Spirochetes.
3. Staining techniques - Simple staining, Gram's staining, Acid fast staining, Meta chromatic granule staining, Spore, Capsule, Flagella.
4. Hematological reactions - Blood Grouping – forward and reverse, Rh Typing
5. Identification of various immune cells by morphology – Leishman staining, Giemsa staining.
6. Agglutination Reactions- Latex Agglutination reactions- RF, ASO, CRP.
7. Detection of HBs Ag by ELISA.
8. Precipitation reactions in gels– Ouchterlony double immunodiffusion (ODD) and Mancini's single radial immunodiffusion (SRID)
9. Immuno-electrophoresis and staining of precipitin lines- Rocket immuno electrophoresis and counter current immuno electrophoresis.
10. Preparation of lymphocytes from peripheral blood by density gradient centrifugation.
11. Purification of immunoglobulin– Ammonium Sulphate Precipitation.

12. Separation of IgG by chromatography using DEAE cellulose or Sephadex.

13. Western Blotting – Demonstration.

14. Isolation of genomic DNA from *E. coli* and analysis by agarose gel electrophoresis

15. Estimation of DNA using colorimeter (Diphenylamine reagent)

16. Separation of proteins by polyacrylamide gel electrophoresis (SDS-PAGE)

17. UV induced mutation and isolation of mutants by replica plating technique.

18. Plasmid DNA isolation from *E.coli*.

19. RNA isolation from yeast.

20. RNA estimation by Orcinol method.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Apply microscopic techniques and staining methods in the identification and differentiation of microbes.	PO1, PO6, PO7, PO8, PO9, PO11
CO2	Apply the knowledge on the sterilization of glass wares and media by different methods and measurement of cell growth.	PO1, PO6, PO7, PO8, PO9, PO11
CO3	Perform and evaluate immunological reactions to aid diagnosis.	PO5, PO7, PO8, PO9, PO11
CO4	Assess the level of lymphocytes in a blood sample and purify immunoglobulin employing appropriate techniques.	PO6, PO7, PO8, PO9, PO11
CO5	Perform DNA extraction and gene transfer mechanisms, analyze and identify by gel electrophoresis	PO6, PO7, PO8, PO9, PO11

Text Books

1. Dubey R.C. and Maheshwari D. K. (2010). Practical Microbiology. S. Chand.
2. Cappuccimo, J. and Sherman, N. (2002). Microbiology: A Laboratory Manual, (6th Edition). Pearson Education, Publication, New Delhi.
3. Cullimore D. R. (2010). Practical Atlas for Bacterial Identification. (2nd Edition). -Taylor &Francis.
4. Rich R. R., Fleisher T. A., Shearer W. T., Schroeder H, Frew A. J. and Weyand C. M. (2018). Clinical Immunology: Principles and Practice. (5th Edition). Elsevier.

5. Glick B. R. and Patten C.L. (2018). Molecular Biotechnology – Principles and Applications of Recombinant DNA. (5th Edition). ASM Press.

Reference Books

1. Collee J. G., Fraser A.G. Marmion B. P. and Simmons A. (1996). Mackie & McCartney Practical Medical Microbiology. (14th Edition). Elsevier, New Delhi.
2. Gupta P. S. (2003). Clinical Immunology. Oxford University Press.
3. Brown T.A. (2016). Gene Cloning and DNA Analysis. (7th Edition). John Wiley and Jones, Ltd.
4. Dale J. W., Schantz M.V. and Plant N. (2012). From Gene to Genomes – Concepts and Applications of DNA Technology. (3rd Edition). John Wileys and Sons Ltd. 2012.
5. Maloy S. R., Cronan J.E. Jr. and Freifelder D. (2011). Microbial Genetics. (2nd Edition). Narosa Publishing Home Pvt Ltd.

Web Resources

1. http://textbookofbacteriology.net/
2. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC149666/
3. https://ocw.mit.edu/courses/hst-176-cellular-and-molecular-immunology-fall-2005/pages/lecture-notes/
4. [PDF] Lehninger Principles of Biochemistry (8th Edition) By David L. Nelson and Michael M. Cox Book Free Download - StudyMaterialz.in

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M					S	M	M	S		M			
CO2	M					S	M	M	S		M			
CO3					S		S	M	S		M			
CO4						S	S	M	S		S			
CO5						S	S	M	S		S			

FORENSIC SCIENCE

Course Code: P1R3MBDSE1:1	**Credit: 3**
Category: Discipline Specific Elective	**Hrs/Week: 5**, **Total Inst.Hrs:** 75
Nature of the Course: Employability	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Understand the Scope, need and learn the tools and techniques in forensic science.
2. Comprehend organizational setup of a forensic science laboratory.
3. Identify and Examine body fluids for identification.
4. Extract DNA from blood samples for investigation.
5. Recognize medico legal post mortem procedures and their importance.

Unit I **15 hrs**

Forensic Science - Definition, history and development of forensic science. Scope and need of forensic science in present scenario. Branches of forensic science. Tools and techniques of forensic science. Duties of a forensic scientist.

Unit II **15 hrs**

Forensic science laboratories - Organizational setup of a forensic science laboratory. Central and State level laboratories in India. Mobile forensic science laboratory and its functions. Forensic microbiology - Types and identification of microbial organisms of forensic significance.

Unit III **15 hrs**

Forensic serology - Definition, identification and examination of body fluids - Blood, semen, saliva, sweat and urine. Forensic examination and identification of hair and fibre.

Unit IV **15 hrs**

DNA profiling - Introduction, history of DNA typing. Extraction of DNA from blood samples - Organic and Inorganic extraction methods. DNA fingerprinting - RFLP, PCR, STR. DNA testing in disputed paternity.

Unit V **15 hrs**

Forensic toxicology - Introduction and concept of forensic toxicology. Medico legal post mortem and their examination. Poisons - Types of poisons and their mode of action.

Course Outcomes	On completion of this course, students will;	
CO1	Identify the scope and need of forensic science in the present scenario.	PO1, PO6, PO7, PO8, PO9
CO2	Plan for the organizational setup and functioning of forensic science laboratories.	PO1, PO6, PO7, PO8, PO9
CO3	Analyze the biological samples found at the crime scene.	PO1, PO5, PO7, PO8, PO9
CO4	Perform extraction and identification of DNA obtained from body fluids.	PO1, PO6, PO7, PO8, PO9
CO5	Discuss the concept of forensic toxicology.	PO1, PO6, PO7, PO8, PO9

Text Books

1. Nanda B. B. and Tewari R. K. (2001) Forensic Science in India: A Vision for the Twenty First Century. Select Publishers, New Delhi. ISBN- 10:8190113526 / ISBN-13:9788190113526.
2. James S. H. and Nordby, J. J. (2015) Forensic Science: An Introduction to Scientific and Investigative Techniques. (5th Edition). CRC Press. ISBN-10:9781439853832 / ISBN-13:978-1439853832.
3. Li R. (2015) Forensic Biology. (2nd Edition). CRC Press, New York. ISBN-13:978-1-4398-8972-5.
4. Sharma B.R (2020) Forensic science in criminal investigation and trials. (6th Edition)Universal Press.
5. Richard Saferstein (2017). Criminalistics- An introduction to Forensic Science. (12th Edition).Pearson Press.

References Books

1. Nordby J. J. (2000). Dead Reckoning. The Art of Forensic Detection- CRC Press, New York. ISBN:0-8493-8122-3.

2. Saferstein R. and Hall A. B. (2020). Forensic Science Hand book, Vol. I, (3rd Edition). CRC Press, New York. ISBN-10:1498720196.

3. Lincoln, P.J. and Thomson, J. (1998). (2nd Edition). Forensic DNA Profiling Protocols. Vol. 98. Humana Press. ISBN: 978-0-89603-443-3.

4. Val McDermid (2014). Forensics. (2nd Edition). ISBN 9780802125156.

5. Vincent J. DiMaio., Dominick DiMaio. (2001). Forensic Pathology (2nd Edition). CRC Press.

Web resources

1. http://clsjournal.ascls.org/content/25/2/114
2. https://www.ncbi.nlm.nih.gov/books/NBK234877/
3. https://www.elsevier.com/books/microbial-forensics/budowle/978-0-12-382006-8
4. https://www.researchgate.net/publication/289542469_Methods_in_microbial_forensics
5. https://cisac.fsi.stanford.edu/events/microbial forensics

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	L					S	M	M	S					
CO2	M					S	M	M	S					
CO3	L				S		S	M	S					
CO4	M					S	S	M	S					
CO5	M					S	S	M	S					

HEALTH AND HYGIENE

Course Code: P1R3MBDSE1:2	**Credit: 3**
Category: Discipline Specific Elective	**Hrs/Week: 5**, **Total Inst.Hrs:** 75
Nature of the Course: Skill Development	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Acquire knowledge on hygiene and live healthy.
2. Provide insights on health laws for food safety and hygiene.
3. Explain health, physical exercises and their importance.
4. Illustrate mental hygiene and involved in mental hygiene.
5. Describe the various health and health education programmes by the government.

Unit I **15 hrs**

Introduction to hygiene and healthful live. Factors affecting health, health habits and practices. Recognizing positive & negative practices in the community. Scientific principles related to health.

Unit II **15 hrs**

Nutrition and Health – Balanced diet, Food surveillance, food Fortification, adulteration and preventive measures. Health laws for food safety. Environmental and housing hygiene. Ventilation and lighting.

Unit III **15 hrs**

Physical health, physical exercises and their importance – Walking, jogging, yoga and meditation, stress relief. International control of health, WHO. Personal hygiene, Sun bathing, Colon Hygiene. Health destroying habits and addictions - Pan, supari, ganja, drinking, smoking, tea and coffee.

Unit IV **15 hrs**

Mental hygiene - factors responsible, developmental tasks, basic needs, emotional stability. Mental hygiene and health in infancy, early childhood, adolescence, adulthood and old age. Mental health occupational hazards.

Unit V **15 hrs**

Health programme and health education – Malaria control, Tuberculosis control, AIDS control programmes and Immunization Programmes. Family planning, Reproductive and Child health programmes (RCH).

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Identify factors affecting health and health habits.	PO1, PO5, PO10
CO2	Execute the knowledge of ventilation and lighting. Justify Health laws for food safety and hygiene.	PO5, PO10
CO3	Follow personal hygiene to avoid diseases and Prevent people from health-destroying habits and addictions.	PO5, PO10
CO4	Explore Mental hygiene and maintain emotional stability.	PO5, PO10
CO5	Participate in health education programmes	PO1, PO5, PO10

Text Books

1. Bamji M. S., Krishnaswamy K. and Brahmam G. N. V. (2019). Textbook of Human Nutrition. (4th Edition). Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi
2. Swaminathan (1995) Food& Nutrition (Vol I) (2nd Edition). The Bangalore Printing &Publishing Co Ltd., Bangalore.
3. Paniker J. C. K. and Ananthanarayan R. (2017). Textbook of Microbiology. (10th Edition). Universities Press (India) Pvt. Ltd
4. Lindsay Dingwall.(2010). Personal Hygiene Care
5. Print ISBN:9781405163071 |Online ISBN:9781444318708 |DOI:10.1002/9781444318708
6. Walter C. C. Pakes(1900). The Science of Hygiene: a Text-book of Laboratory Practice.
7. (London: Methuen and Co.,).

References Books

1. Khader V. (2000) Food, Nutrition and Health, Kalyan Publishers, New Delhi.

2. Srilakshmi, B. (2010) Food Science, (5th Edition) New Age International Ltd., New Delhi.
3. Dubey R.C. and Maheshwari D. K. (2010). Practical Microbiology. S. Chand.
4. Park K. 2007, Park's text book of Preventive and Social Medicine, Banarsidas Bhanot publishers, India.
5. Srilakshmi, 2002, Dietetics, New Age Publications, India

Web Resources

1. Health and Hygiene - Personal Hygiene, Community Hygiene and Diseases (vedantu.com)
2. Chapter-32.pdf (nios.ac.in)
3. Menstrual Health and Hygiene Guide | Student Health and Counseling Services (ucdavis.edu)
4. https://nap.nationalacademies.org/read/11756/chapter/13
5. http://ecoursesonline.iasri.res.in/mod/page/view.php?id=112325

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	L				S					M				
CO2					S					M				
CO3					S					L				
CO4					S					M				
CO5	L				S					M				

MICROALGAL TECHNOLOGY

Course Code: P1R3MBDSE1:3	**Credit: 3**
Category: Discipline Specific Elective	**Hrs/Week: 5**, **Total Inst.Hrs:** 75
Nature of the Course: Skill Development	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Characterize the different groups of algae.
2. Describe the cultivation and harvesting of algae.
3. Identify the commercial applications of various algal products.
4. Apply microalgae for environmental applications.
5. Employ microalgae as alternate fuels.

Unit I **15 hrs**

Introduction to Algae - General characteristics. Classification of algae according to Fritsch. Salient features of different groups of algae. Distribution - Freshwater, brackish water and marine algae. Identification methods. An overview of applied Phycology. Economically important microalgae.

Unit II **15 hrs**

Cultivation of freshwater and marine microalgae - Growth media. Isolation and enumeration of microalgae. Laboratory cultivation and maintenance. Outdoor cultivation - Photobioreactors - construction, types and operation; raceway ponds - Heterotrophic and mixotrophic cultivation - Harvesting of microalgae biomass.

Unit III **15 hrs**

Microalgae in food and nutraceutical applications - Algal single cell proteins. Cultivation of Spirulina and Dunaliella. Microalgae as aquatic, poultry and cattle feed. Microalgal biofertilizers. Value-added products from microalgae. Pigments - Production of microalgal carotenoids and their uses. Phycobiliproteins - production and commercial applications. Polyunsaturated fatty acids as active nutraceuticals. Microalgal secondary metabolites - Pharmaceutical and cosmetic applications.

Unit IV **15 hrs**

Microalgae in environmental applications. Phycoremediation - Domestic and industrial waste water treatment. High-rate algal ponds and surface-immobilized systems - Treatment of gaseous wastes by microalgae. Sequestration of carbon dioxide. Scavenging of heavy metals by microalgae. Negative effects of algae. Algal blooms, algicides for algal control.

Unit V **15 hrs**

Microalgae as feed stock for production of biofuels - Carbon-neutral fuels. Lipid-rich algal strains - Botryococcus braunii. Drop-in fuels from algae - hydrocarbons and biodiesel, bioethanol, biomethane, biohydrogen and syngas from microalgae biomass. Biocrude synthesis from microalgae. Integrated biorefinery concept. Life cycle analysis of algae biofuels.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Acquire knowledge in the field of microalgal technology and their characteristics.	PO1
CO2	Identify the methods of algal cultivation and harvesting.	PO1, PO6
CO3	Recognize and recommend the use of microalgae as food, feed and fodder.	PO7, PO8, PO9
CO4	Promote microalgae in phycoremediation.	PO7, PO9, PO11, PO14
CO5	Compare and critically evaluate recent applied research in these microalgal applications.	PO7, PO8, PO9

Text Books

1. Lee R.E. (2008). Phycology. Cambridge University Press.
2. Sharma O.P. (2011). Algae. Tata McGraw-Hill Education.
3. Shekh A., Schenk P., Sarada R. (2021). Microalgal Biotechnology. Recent Advances, Market Potential and Sustainability. Royal Society of Chemistry.
4. Lele. S.S., Jyothi Kishen Kumar (2008). Algal bio process technology. New Age International P(Ltd)
5. Das., Mihirkumar. Algal Biotechnology. Daya Publishing House, New Delhi.

References Books

1. Andersen R.A. (2005). Algal culturing techniques. Academic Press, Elsevier.

2. Bux F. (2013). Biotechnological Applications of Microalgae: Biodiesel and Value-added Products. CRC Press.
3. Singh B., Bauddh K., Bux, F. (2015). Algae and Environmental Sustainability. Springer.
4. Das D. (2015). An algal biorefinery: An integrated approach. Springer.
5. Bux F. and Chisti Y. (2016). Algae Biotechnology: Products and Processes. Springer.

Web Resources

1. https://www.classcentral.com/course/algae-10442
2. https://onlinecourses.nptel.ac.in/noc19_bt16/preview
3. https://freevideolectures.com/course/4678/nptel-industrial-biotechnology/46
4. https://nptel.ac.in/courses/103103207
5. https://www.sciencedirect.com/topics/earth-and-planetary-sciences/microalgae

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S													
CO2	S					M								
CO3							S	S	S					
CO4							S		S		M			M
CO5							M	S	S					

BIOINSTRUMENTATION

Course Code: P1R3MBDSE2:1	**Credit: 3**
Category: Discipline Specific Elective	**Hrs/Week: 5**, **Total Inst.Hrs:** 75
Nature of the Course: Employability	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Explain the principles and working mechanisms of laboratory instruments.
2. Discuss chromatography techniques and molecular biology techniques.
3. Illustrate molecular techniques in biological applications.
4. Acquire knowledge on spectroscopic techniques
5. Demonstrate the use of radio isotopes in various techniques.

Unit I **15 hrs**

Basic laboratory Instruments. Aerobic and anaerobic incubator – Biosafety Cabinets - Fume Hood, pH meter, Lyophilizer, Flow cytometry. Centrifugation techniques: Basic principles of centrifugation - Standard sedimentation coefficient - measurement of sedimentation co-efficient; Principles, methodology and applications of differential, rate zonal and density gradient centrifugation - Applications in determination of molecular weight.

Unit II **15 hrs**

General principles of chromatography - Chromatographic Performance parameters; Types- Thin layer chromatography, Paper Chromatography, Liquid chromatography (LPLC &HPLC), Adsorption, ion exchange, Gel filtration, affinity, Gas liquid (GLC). Flash Chromatography and Ultra Performance convergence chromatography. Two dimensional chromatography. Stimulated moving bed chromatography (SEC).

Unit III **15 hrs**

Electrophoresis: General principles - moving boundary electrophoresis - electrophoretic mobility – supportive materials – electro endosmosis – types (horizontal, vertical and two dimensional electrophoresis) - Principle and applications - paper electrophoresis, Serum electrophoresis, starch gel electrophoresis, Disc gel, Agarose gel, SDS – PAGE, Immuno electrophoresis. Blotting techniques -Southern, northern and western blotting.

Unit IV **15 hrs**

Spectroscopic techniques: Principle, simple theory of absorption of light by molecules, electromagnetic spectrum, instrumentation and application of UV- visible, Raman, FTIR spectrophotometer, spectrofluorimetry, Atomic Absorption Spectrophotometer, Flame spectrophotometer, NMR, ESR, Emission Flame Photometry and GC-MS. Detection of molecules in living cells - FISH and GISH. Biophysical methods: Analysis of biomolecules by Spectroscopy UV/visible.

Unit V **15 hrs**

Radioisotopic techniques: Principle and applications of tracer techniques in biology. Radioactive isotopes - radioactive decay; Detection and measurement of radioactivity using ionization chamber, proportional chamber, Geiger- Muller and Scintillation counters, auto radiography and its applications. Commonly used isotopes in biology, labeling procedures and safety aspects.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Make use of the laboratory instruments- laminar air flow, pH meter, centrifugation methods, biosafety cabinets following SOP.	PO4, PO6, PO7, PO8, P11
CO2	Apply chromatography techniques in the separation of biomolecules.	PO4, PO6, PO7, PO8, P11
CO3	Perform molecular techniques like mutagenesis and their detection.	PO4, PO6, PO7, PO8, P11
CO4	Estimate molecules in biological samples by adopting UV spectroscopic techniques.	PO4, PO6, PO7, PO8, P11
CO5	Cultivate organisms anaerobically.	PO4, PO6, PO7, PO8, P11

Text Books

1. Sharma B. K. (2014). Instrumental Method of Chemical Analysis. Krishna Prakashan Media (P) Ltd.
2. Chatwal G. R and Anand S. K. (2014.) Instrumental Methods of Chemical Analysis. Himalaya Publishing House.
3. Mitchell G. H. (2017). Gel Electrophoresis: Types, Applications and Research. Nova Science Publishers Inc.

4. Holme D. Peck H. (1998). Analytical Biochemistry. (3rd Edition). Prentice Hall.
5. Jayaraman J. (2011). Laboratory Manual in Biochemistry. (2ndEdition). Wiley Eastrn Ltd., New Delhi.

References Books

1. Pavia D. L. (2012) Spectroscopy (4th Edition). Cengage.
2. Skoog A. and West M. (2014). Principles of Instrumental Analysis. (14th Edition). W.B.Saunders Co., Philadephia.
3. Miller J. M. (2007). Chromatography: Concepts and Contrasts (2nd Edition) Wiley-Blackwell.
4. Gurumani N. (2006). Research Methodology for Biological Sciences. (1st Edition) MJP Publishers.
5. Ponmurugan P. and Gangathara P. B. (2012). Biotechniques. (1st Edition). MJP Publishers.

Web Resources

1. https://norcaloa.com/BMIA
2. http://www.biologydiscussion.com/biochemistry/centrifugation/centrifuge-introduction-types-uses-and-other-details-with-diagram/12489
3. https://www.watelectrical.com/biosensors-types-its-working-and-applications.
4. http://www.wikiscales.com/articles/electronic-analytical-balance/
5. https://study.com/academy/lesson/what-is-chromatography-definition-types-uses.

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1				S		M	M	S			S			
CO2				S		M	M	S			S			
CO3				S		S	S	S			S			
CO4				S		M	S	S			S			
CO5				S		M	S	S			L			

HERBAL TECHNOLOGY AND COSMETIC MICROBIOLOGY

Course Code: P1R3MBDSE2:2	**Credit: 3**
Category: Discipline Specific Elective	**Hrs/Week: 5**, **Total Inst.Hrs:** 75
Nature of the Course: Employability	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Impart knowledge of Indian Medicinal Plants and their applications in microbiology.
2. Promote the technical skills involved in preparation of different types of plant extracts.
3. Explain methods to analyze the antimicrobial activity of medicinal plants.
4. Acquire knowledge on cosmetic microbiology and role of microorganisms in cosmetics.
5. Gain insight into pharmacopeial microbial assays and biosafety.

Unit I **15 hrs**

Herbs, Herbal medicine - Indian medicinal plants: Scope and Applications of Indian medicinal plants in treating bacterial, fungal and viral diseases. Basic principles involved in Ayurvedha, Sidha, Unani and Homeopathy.

Unit II **15 hrs**

Collection and authentication of selected Indian medicinal plants: *Emblica officinalis, Withania somnifera, Phyllanthus amarus, Tinospora cordifolia, Andrographis paniculata, Piper longum, Ocimum sanctum, Azardirchata indica, Terminalia chebula, Allium sativum*. Preparation of extracts- Hot and cold methods. Preparation of stock solutions.

Unit III **15 hrs**

Antimicrobial activity of selected Indian medicinal Plants: - In vitro determination of antibacterial and fungal activity of selected whole medicinal plants/ parts – well-diffusion methods. MIC - Macro and micro dilution techniques. Antiviral activity- cell lines- cytotoxicity, cytopathic and non-cytopathic effect.

Unit IV **15 hrs**

History of Cosmetic Microbiology – Need for cosmetic microbiology, Scope of cosmetic microbiology, - Role of microbes in cosmetic preparation. Preservation of cosmetics. Antimicrobial properties of natural cosmetic products – Garlic, neem, turmeric, aloe vera and tulsi. Sanitary practices in cosmetic manufacturing - HACCP protocols in cosmetic microbiology.

Unit V **15 hrs**

Cosmetic microbiology test methods - Antimicrobial preservative efficacy, microbial content testing and biological toxicological testing. Validation methods - bioburden and Pharmacopeial microbial assays. Preservatives of cosmetics - Global regulatory and toxicological aspect of cosmetic preservatives.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Identify the applications of Indian medicinal plants in treating diseases.	PO1, PO5
CO2	Identify and authenticate herbal plants.	PO6, PO7
CO3	Evaluate the antimicrobial activity of medicinal plants.	PO4, PO6, PO9
CO4	Describe the role of microorganisms and their metabolites in the preparation of cosmetics.	PO1, PO5, PO7
CO5	Validate procedures and biosafety measures in the mass production of cosmetics.	PO6, PO7

Text Books

1. Ayurvedic Formulary of India. (2011). Part 1, 2 & 3. Pharmacopoeia Commission for Indian Medicine and Homeopathy. ISBN-10:8190648977.
2. Panda H. (2004). Handbook on herbal medicines. Asia Pacific Business Press Inc. ISBN:8178330911.

3. Mehra P. S. (2019). A Textbook of Pharmaceutical Microbiology. Dreamtech Press. ISBN 13:9789389307344.
4. Geis P. A. (2020). Cosmetic microbiology: A Practical Approach. (3rd Edition). CRC Press. ISBN:9780429113697.
5. Brannan D. K. (1997). Cosmetic microbiology: A Practical Handbook. CRC Press.ISBN-10:0849337135.

References Books

1.Indian Herbal Pharmacopoeia (2002). Vol. I &II Indian Drug Manufacturers Association, Mumbai.
2.British Herbal Pharmacopoeia.(1990).Vol.I. British Herbal Medicine Association.ISBN: 0903032090.
3.Verpoorte R. and Mukherjee, P. K. (2010). GMP for Botanicals: Regulatory and Quality issues on Phytomedicines. In GMP for botanicals: regulatory and quality issues on phytomedicines. (2nd edition). Saujanya Books, Delhi.ISBN-10:81-900788-5-2/8190078852. ISBN-13:978-81-900788-5-6/9788190078856.
4.Turner R. (2013). Screening methods in Pharmacology. Elsevier. ISBN:9781483264233.
5.Cupp M. J. (2010). Toxicology and Clinical Pharmacology of Herbal Products (pp. 85-93). M. J. Cupp. Humana Press.Totowa, NJ, USA. ISBN-10:1617371904.

Web Resources

1. https://www.academia.edu/50236711/Modern_Extraction_Methods_for_Preparation_of_Bioactive_Plant_Extracts
2. https://www.nhp.gov.in/introduction-and-importance-of-medicinal-plants-and-herbs_mtl

3. https://pubmed.ncbi.nlm.nih.gov/17004305/
4. https://www.fda.gov/cosmetics/potential-contaminants-cosmetics/microbiological-safety-and-cosmetics
5. https://pubmed.ncbi.nlm.nih.gov/15156038/

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M				S									
CO2						S	M							
CO3				S		S			M					
CO4	M				S		S							
CO5						M	S							

ESSENTIALS OF LABORATORY MANAGEMENT AND BIOSAFETY

Course Code: P1R3MBDSE2:3	**Credit: 3**
Category: Discipline Specific Elective	**Hrs/Week: 5**, **Total Inst.Hrs:** 75
Nature of the Course: Employability	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. To utilize containment principles to ensure biosafety.
2. To enrich the student role and responsibilities of laboratory hazards and their control.
3. To know the importance of first aid technique for various common lab accidents.
4. To acquire knowledge of biosafety level, risk assessment and maintain proper hygiene in the laboratory.
5. To discuss the biosafety regulations and guidelines and implementation of safety programs.

Unit I **15 hrs**

Introduction to the laboratory and laboratory hazards - General laboratory facilities – Occupational safety- Lab accidents - Fires, chemical burns, slips and falls, Animal bites. Cuts from broken glass. Toxic fume inhalation. General laboratory rules, Good laboratory practice (GLP). Laboratory plan.

Unit II **15 hrs**

Common hazards in laboratory: Chemical hazards- Safe handling of chemicals and gases, hazard labels and symbols. Material safety datasheet (MSDS), Chemical handling - Fume hood, Storage of chemicals. Chemical Waste Disposal Guideline. Physical hazards - Physical agent data sheets (PADS), Electric hazards- Electrical shock, Electrical explosions, Electrical burns. Safe work practices. Potential ignition sources in the lab. Stages of Fire. Fire Extinguishers. Fire Response.

Unit III **15 hrs**

Prevention and First aid for laboratory accidents. Personal protective equipment (PPE), Proper attire (Eye/Face Protection, laboratory coats, gloves, respirators. Disposal/Removal of PPE.

Emergency equipment safety - Showers/ Eye Washes. Laboratory security and emergency response. First aid for - Injuries caused by broken glass, Acid/Alkali splashes on the skin, swallowing acid/alkali, burns caused by heat, electric shock.

Unit IV **15 hrs**

Biosafety - Historical background. Blood borne pathogens (BBP) and laboratory - acquired infections. Introduction to biological safety cabinets. Primary containment for biohazards. Biosafety levels of specific microorganisms. Recommended biosafety. Levels for infectious agents and infected animals. Risk groups with examples - Risk assessment. Safety levels. Case studies - Safe working, hand hygiene. Laboratory instruments, packing, sending, transport, import and export of biological agents. Hygiene, disinfection, decontamination, sterilization.

Unit V **15 hrs**

Biosafety regulations and guidelines. Centers for disease control and prevention and the National institutes of health. Occupational safety and health administration. Recombinant DNA advisory committee(RDAC), Institutional biosafety committee(IBSC), Review committee on genetic manipulation(RCGM), Genetic engineering approval committee (GEAC). Implementation of biosafety guidelines.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Employ skills on laboratory safety and avoid laboratory accidents.	PO1, PO2, PO3, PO7, PO11
CO2	Prevent laboratory hazards by practicing safety strategies.	PO2, PO5, PO7, PO11
CO3	Practice various first aid procedures during common laboratory accidents.	PO1, PO2, PO3, PO5, PO10, PO11
CO4	Ensure biosafety strategies in laboratory.	PO2, PO3, PO4, PO7, PO10, PO11
CO5	Recognize the importance of biosafety guidelines.	PO3, PO4, PO5, PO7, PO10, PO11

Text Books

1. Sateesh M. K. (2013). Bioethics and Biosafety, IK International Pvt Ltd. ISBN : 8190675702.
2. Muthuraj M. and Usharani B. (2019). Biosafety in Microbiological Laboratories. (1sr Edition). Notion Press. ISBN 10: 1645878856
3. Biosafety in Microbiological and Biomedical Laboratories - U.S. Health Department and Human Services. (2016). (5th Edition). Lulu.com.
4. Kanai. L. Mukherjee. (Medical Laboratory Technology(4th Edition). CBS Publishers.
5. Ramakrishnan (2012). Manual of Medical Laboratory Techniques. JP brothers.

References Books

1. World Health Organization, Biosafety programme management. (2010). (4th Edition). WHO Publications.
2. Rashid N. (2013). Manual of Laboratory Safety (Chemical, Radioactive, and Biosafety with Biocides) (1st Edition).
3. Dayuan X. (2015). Biosafety and Regulation for Genetically Modified Organisms, Alpha Science International Ltd, ISBN-10 1842657917:
4. Ochei J. Kolhatkar(2000). A. (Medical Laboratory Science – Theory and Practice. ISBN; 13:978-0074632239.
5. Lynne S. Garcia. Clinical Laboratory Management (2nd Edition). ASM Press

Web Resources

1. https://www.cdc.gov/labs/pdf/CDC-
2. iosafetymicrobiologicalBiomedicalLaboratories-2009-P.pdf
3. https://ucanapplym.s3.ap-south-1.amazonaws.com/RGU/notifications/E_learning/0nline_study/PG-SEM-IV-Biosafety%20regulation.pdf
4. https://consteril.com/biosafety-levels-difference/
5. https://www.cdc.gov/labs/pdf/CDCBiosafetymicrobiologicalBiomedicalLaboratories-2009-P.pdf

6. https://www.who.int/publications/i/item/9789240011311

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S	S	S				S				S			
CO2		S			S		S				S			
CO3	S	S	S		S					S	S			
CO4		S	S	M			S			S	S			
CO5			S	S	S		S			S	S			

SEMESTER II

MEDICAL BACTERIOLOGY AND MYCOLOGY

Course Code: P2R3MBCC4	**Credit:** 5
Category: Core Course	**Hrs/Week:** 6, **Total Inst.Hrs:** 90
Nature of the Course: Skill Development	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Acquire Knowledge on collection, transportation and processing of various kinds of clinical specimens.
2. Explain morphology, characteristics and pathogenesis of bacteria.
3. Discuss various pathogenesis of enterobacteria.
4. Acquire knowledge on antifungal agents and their importance.
5. Describe various diagnostic methods available for fungal disease diagnosis.

Unit I: **20 hrs**

Classification of medically important bacteria, Normal flora of human body, Collection, transport, storage and processing of clinical specimens, Microbiological examination of clinical specimens, antimicrobial susceptibility testing. Handling and maintenance of laboratory animals – Rabbits, guinea pigs and mice.

Unit II: **20 hrs**

Morphology, classification, characteristics, pathogenesis, laboratory diagnosis and treatment of diseases caused by species of *Staphylococci, Streptococci, Pneumococci, Neisseriae., Bacillus, Corynebacteria, Mycobacteria* and *Clostridium.*

Unit III: **20 hrs**

Morphology, classification, characteristics, pathogenesis, laboratory diagnosis and treatment of diseases caused by Enterobacteriaceae members, *Yersinia, Pseudomonas, Vibrio, Mycoplasma, Helicobacter, Rickettsiae, Chlamydiae, Bordetella, Francisella., Spirochaetes- Leptospira, Treponema* and *Borrelia.* Nosocomial, zoonotic and opportunistic infections -prevention and control.

Unit IV: **15 hrs**

Morphology, taxonomy and classification of fungi, detection and recovery of fungi from clinical specimens. Dermatophytes and agents of superficial mycoses. *Trichophyton, Epidermophyton & Microsporum*. Yeasts of medical importance – *Candida, Cryptococcus*. Mycotoxins, Antifungal agents, testing methods and quality control.

Unit V: **15 hrs**

Dimorphic fungi causing Systemic mycoses, *Histoplasma cocciodioides, Sporothrix, Blastomyces*. Fungi causing Eumycotic Mycetoma, Opportunistic fungi- Newer methods in diagnostic mycology.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Collect, transport and process of various kinds of clinical specimens.	PO1,PO5,PO9
CO2	Analyze various bacteria based on morphology and pathogenesis.	PO1,PO5,PO9
CO3	Discuss various treatment methods for bacterial disease.	PO1,PO5,PO9
CO4	Employ various methods detect fungi in clinical samples and apply knowledge on antifungal agents..	PO5,PO9
CO5	Apply various immunodiagnostic method to detect fungal infections.	PO5,PO9

Text Books

1. Kanunga R. (2017). Ananthanarayanan and Panicker's Text book of Microbiology. (2017).Orient Longman, Hyderabad.
2. Greenwood, D., Slack, R. B. and Peutherer, J. F. (2012) Medical Microbiology, (18th Edition). Churchill Livingstone, London.
3. Finegold, S. M. (2000) Diagnostic Microbiology, (10th Edition). C.V. Mosby Company, St. Louis.
4. Alexopoulos C. J., Mims C. W. and Blackwell M. (2007). Introductory Mycology, (4th Edition). Wiley Publishers.
5. Chander J. (2018). Textbook of Medical Mycology. (4th Edition). Jaypee brothers Medical Publishers.

Reference Books

1. Salle A. J. (2007). Fundamental Principles of Bacteriology. (4th Edition). Tata McGraw-Hill Publications.
2. Collee J.C. Duguid J.P. Foraser, A.C, Marimon B.P, (1996). Mackie & McCartney Practical Medical Microbiology. 14thedn, Churchill Livingston.
3. Cheesbrough M. (2006). District Laboratory Practice in Tropical countries.- Part 22ndedn.Cambridge University Press.
4. Topley and Wilson's. (1998). Principles of Bacteriology.9th edn. Edward Arnold, London.
5. Murray P.R., Rosenthal K.S. and Michael A. (2013). Medical Microbiology. Pfaller. 7th edn. Elsevier, Mosby Saunders.

Web Resources

1. http://textbookofbacteriology.net/nd
2. https://microbiologysociety.org/members-outreach-resources/links.html
3. https://www.pathelective.com/micro-resources
4. http://mycology.cornell.edu/fteach.html
5. https://www.adelaide.edu.au/mycology/

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M				S				M					
CO2	M				S				M					
CO3	M				S				M					
CO4					S				M					
CO5					S				M					

MEDICAL VIROLOGY AND PARASITOLOGY

Course Code: P2R3MBCC5	**Credit:** 5
Category: Core Course	**Hrs/Week:** 6, **Total Inst.Hrs:** 90
Nature of the Course: Skill Development	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Describe the classification and identification of viruses.
2. Acquire knowledge about the life cycle of viruses.
3. Develop diagnostic and pathogenesis of Animal viruses.
4. Impart knowledge about parasitic infections.
5. Develop diagnostic skills, in the identification of parasitic infections.

Unit I: **20 hrs**

Historical perspectives- Morphology of viruses-General properties of viruses-Current ICTV Classification of viruses, Cultivation of viruses-Identification of virus-infected cells, sub viral agents- viroids, virusoids, and prions. Serodiagnosis

Unit II: **20 hrs**

Life cycle of Bacteriophages: lytic and lysogenic cycle- One step growth curve. General life cycle of bacteriophages: Øx174, M13, Mu, T4, λ, and P1 phages. Phage typing.

Unit III: **15 hrs**

Laboratory diagnosis and pathogenesis of Animal viruses: DNA viruses - Adeno, Pox, Herpes and Hepadna viruses. RNA viruses- Polio, Rabies and Retroviruses. Arboviruses: Dengue & Chikungunya, Ebola, H1N1 virus. Other emerging viral diseases. Viral vaccines and drugs.

Unit IV: **15 hrs**

Introduction to Medical Parasitology – Classification, Laboratory Techniques in Parasitology. Intestinal amoeba – *Entamoeba hystolytica*, Free living amoeba – *Maeglaria fowleri*,

Acanthamoeba Sp. Intestinal and genital flagellate – *Giardia lamblia, Trichomonas vaginalis.* Blood and tissue flagellates, *Leishmania donovani, Trypanosoma cruzi, and Trypanosoma brucei. Malarial parasites, Toxoplasma gondii, Cryptosporidium* Sp.

Unit V: **20 hrs**

Infection of Helminthes - Cestodes – *Taenia solium, T. saginata, Echinococcus granulosus.* Trematodes – F*asciola hepatica, Paragonimus westermani and Schistosomes*. Nematodes – *Ascaris lumbricoides, Ancylostoma duodenale, Trichuris trichiura, Enterobius vermicularis, Wuchereria bancrofti*.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Cultivate viruses by different methods and aid in diagnosis. Perform purification and viral assay.	PO5, PO7, PO8, PO10
CO2	Investigate the symptoms of viral infections and presumptively identify the viral disease.	PO5, PO7, PO8, PO10
CO3	Diagnose various viral diseases by different methods.(serological, conventional and molecular)	PO5, PO7, PO8, PO10
CO4	Educate public about the spread, control and prevention of parasitic diseases.	PO5, PO7, PO8, PO10
CO5	Identify the protozoans and helminthes present in stool and blood specimens. Perform serological and molecular diagnosis of parasitic infections.	PO5, PO7, PO8, PO10

Text Books

1. Kanunga R. (2017). Ananthanarayanan and Panicker's Text book of Microbiology. (10th Edition). Universities Press (India) Pvt. Ltd.
2. Dubey, R.C. and Maheshwari D.K. (2010). A Text Book of Microbiology. S. Chand & Co.
3. Rajan S. (2007). Medical Microbiology. MJP publisher.
4. Paniker J. (2006). Text Book of Parasitology. Jay Pee Brothers, New Delhi.
5. Arora, D. R. and Arora B. B. (2020). Medical Parasitology. (5th Edition). CBS Publishers & Distributors Pvt. Ltd. New Delhi.

Reference Books

1. Carter J. (2001). Virology: Principles and Applications (1st Edition). Wiley Publications.
2. Willey J., Sandman K. and Wood D. Prescott's Microbiology. (11th Edition). McGraw Hill Book.

3. Jawetz E., Melnick J. L. and Adelberg E. A. (2000). Review of Medical Microbiology. (19th Edition). Lange Medical Publications, U.S.A.
4. Finegold S.M. (2000). Diagnostic Microbiology. (10th Edition). C.V. Mosby Company, St. Louis.
5.

Web Resources

1. https://en.wikipedia.org/wiki/Virology
2. https://academic.oup.com/femsre/article/30/3/321/546048
3. https://www.sciencedirect.com/science/article/pii/S0042682215000859
4. https://nptel.ac.in/courses/102/103/102103039/
5. https://www.healthline.com/health/viral-diseases#contagiousness

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1					M		L	L		M				
CO2					M		L	L		M				
CO3					M		L	L		M				
CO4					M		L	L		M				
CO5					M		L	L		M				

PRACTICAL II

MEDICAL BACTERIOLOGY AND MYCOLOGY & MEDICAL VIROLOGY AND PARASITOLOGY

Course Code: P2R3MBCC6P	**Credit:** 4
Category: Core Course – Practical II	**Hrs/Week:** 6
Nature of the Course: Skill Development	**Marks:** CIA: 40+ EXT: 60 = 100

Course Objectives

1. Develop skills in the diagnosis of bacterial infections and antimicrobial sensitivity.
2. Impart knowledge on fungal infections and identification.
3. Diagnose malarial parasites.
4. To gain knowledge about phages.
5. Screen and utilize the cultivation of virus.

1. Staining of Clinical Specimens – wet mount, differential and special staining methods.
2. Isolation and identification of bacterial pathogens from clinical specimens (Urine, Pus, Sputum, faeces) - cultivation in basal, differential, enriched, selective and special media – Biochemical identification tests.
3. Enumeration of bacteria in urine to detect significant bacteriuria.
4. Antimicrobial sensitivity testing - Kirby Bauer method.
5. Minimum inhibitory concentration (MIC) test.
6. Minimum bactericidal concentration (MBC) test.
7. Examination of different fungi by Lactophenol Cotton blue staining.
8. Examination of different fungi by KOH staining.
9. Identification of Dermatophytes.
10. Isolation of phages from natural resources.
11. Titration of phages.
12. Egg inoculation of viruses – Different routes – Allantoic, amniotic, yolk sac and CAM (Demo).
13. Preparation of Chick embryo fibroblast culture (Demo).
14. Observation of viral inclusion bodies (Slides) – Demo.

15. Normal saline / Lugol's Iodine preparation for the examination of parasites in stool.

16. Examination of faeces by concentration methods

17. Thin and thick blood smears examination for Malariral parasite.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Collection of different clinical samples, transport, culture and examination.	PO7, PO8, PO9
CO2	Identify medically important bacteria, fungus and parasites from the clinical samples by staining and biochemical tests.	PO7, PO8, PO9
CO3	Promote diagnostic skills; interpret laboratory tests in the diagnosis of infectious diseases.	PO7, PO8, PO9, PO10
CO4	Perform antibiotic sensitivity tests and compare with the standard tests.	PO7, PO8, PO9, PO10
CO5	Screening of industrially important microbes for metabolite production.	PO7, PO8, PO9

Text Books

1. Cullimore D. R. (2010). Practical Atlas for Bacterial Identification, 2nd Edition. Publisher-Taylor and Francis.
2. Abbott A.C. (2010). The Principles of Bacteriology. Nabu Press.
3. Parija S. C. (2012). Textbook of Practical Microbiology. Ahuja Publishing House.
4. Cappuccimo, J. and Sherman, N. (2002) Microbiology: A Laboratory Manual, (6thEdition). Pearson Education, Publication, New Delhi.
5. Morag C. and Timbury M.C. (1994)._Medical Virology. 4th Edition. Blackwell Scientific Publishers.

Reference Books

1. Collee J. G., Fraser A.G. Marmion B. P. and Simmons A. (1996). Mackie & McCartney Practical Medical Microbiology. (14th Edition). Elsevier, New Delhi.
2. Chart H. (2018). Practical Laboratory Bacteriology. CRC Press.
3. Moore V. A. (2017). Laboratory Directions for Beginners in Bacteriology. Triste Publishing Ltd.

4. .Cheesbrough M. (2006). District Laboratory Practice in Tropical countries.- Part 22nd Edition. Cambridge University Press.
5. Murray P.R., Rosenthal K.S. and Michael A. (2013). Medical Microbiology. Pfaller. 7th Edition. Elsevier, Mosby Saunders

Web Resources

http://textbookofbacteriology.net/

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7173454/

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3768729/

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC149666/

https://www.intechopen.com/books/current-issues-in-molecular-virology-viral-genetics- and-biotechnological-applications/vaccines-and-antiviral-agents

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1							M	M	M					
CO2							M	M	M					
CO3							M	M	L	L				
CO4							M	M	M	L				
CO5							M	M	M					

EPIDEMIOLOGY

Course Code: P2R3MBDSE3:1	**Credit:** 3
Category: Discipline Specific Elective	**Hrs/Week:** 4, **Total Inst.Hrs:** 60
Nature of the Course: Employability	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Describe the role of epidemiology in public health.
2. Explain about epidemiology tools and disease surveillance methods.
3. Analyze various communicable and non-communicable diseases in India.
4. Discuss on mechanism of antimicrobial resistance.
5. Outline on National health programmes that have been designed to address the issues.

Unit I: **12 Hrs**

Fundamentals of epidemiology - Definitions of epidemiology – Epidemiology of infectious diseases in Public Health. Natural history of disease - Historical aspects of epidemiology. Common risk factors - Epidemiologic Triad - Agent factors, host factors and environmental factors. Transmission basics - Chain of infection, portal of entry. Modes of transmission -Direct and indirect. Stages of infectious diseases. Agents and vectors of communicable diseases of public health importance and dynamics of disease transmission. Epidemiology of Zoonosis - Factors, routes of transmission of bacterial, viral, parasitic and fungal zoonotic agents. Control of zoonosis.

Unit II: **12 Hrs**

Tools of Epidemiology - Measures of Disease - Prevalence, incidence. Index case. Risk rates. Descriptive Epidemiology - Cohort studies, measuring infectivity, survey methodology including census procedures. Surveillance strategies - Disease surveillance, geographical indication system, outbreak investigation in public health and contact investigation.

Unit III: **12 Hrs**

Epidemiological aspects of diseases of national importance - Background to communicable and non-communicable diseases. Vector borne diseases in India. Diarrhoeal diseases. Zoonoses.

Viral haemorrhagic fevers. Mycobacterial infections. Sexually transmitted diseases. Human Immunodeficiency Virus/Acquired Immunodeficiency Syndrome (HIV/AIDS). Emerging disease threats - Severe Acute Respiratory Syndrome (SARS), Covid-19, Ebola, MDR-TB, Malaria, Mucor mycosis, Avian flu. Dengue, Swine Flu, Chikungunya. Epidemiology, prevention, and control of non-communicable diseases - Asthma, Coronary heart disease, Malignancy, diabetes mellitus, respiratory diseases, eye diseases, Dental disorders. Emerging and Re-emerging Diseases.

Unit IV: **12 Hrs**

Mechanisms of Antimicrobial resistance - Multidrug Efflux pumps, Extended Spectrum β-lactamases (ESBL). Hospital acquired infections - Factors, infection sites, mechanisms, Role of Multidrug resistant pathogens. Role of *Pseudomonas, Acinetobacter, Clostridium difficile*, HBV, HCV, Rotavirus, *Cryptosporidium* and *Aspergillus* in Nosocomial infections. Prevention and management of nosocomial infections.

Unit V: **12 Hrs**

National Programmes related to Communicable and Non-Communicable diseases - National Malaria Eradication Programme, Revised National Tuberculosis Control Programme, Vector Borne Disease Control Programme, National AIDS Control Programme, National Cancer Control Programme and National Diabetes Control Programme. Biochemical and immunological tools in epidemiology - Biotyping, Serotyping, Phage typing, FAME (Fatty acid methyl ester analysis), Curie Point PyMS (Pyrolysis Mass spectrometry), Protein profiling, Molecular typing methods.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Apply the knowledge acquired on concepts of epidemiology to clinical and public health environment.	PO1
CO2	Plan various strategies to trace the epidemiology.	PO4, PO5, PO6
CO3	Plan the control of communicable and non-communicable diseases.	PO1, PO5,
CO4	Analyze the implications of drug resistance in the society and design the control of antimicrobial resistance and its management.	PO5,
CO5	Employ National control programs related to Communicable and Non-Communicable diseases with the public.	PO4, PO5,

Text Books

1.Dicker R., Coronado F., Koo. D. and Parrish. R. G. (2012). Principles of Epidemiology in Public Health Practice., (3rd Edition). CDC.

2.Gerstman B. (2013). Epidemiology Kept Simple: An Introduction to Classic and Modern Epidemiology. (3rd Edition). Wiley Blackwell.

3.Greenwood, D., Slack, R. B. and Peutherer, J. F. (2012) Medical Microbiology, (18th Edition). Churchill Livingstone, London.

4.Jawetz E., Melnick J. L. and Adelberg E. A. (2000). Review of Medical Microbiology. (19th Edition). Lange Medical Publications, U.S.A.

5.Dimmok N. J. and Primrose S. B. (1994). Introduction to Modern Virology.5th edn. Blackwell Scientific Publishers.

References Books

1.Bhopal R. S. (2016).Concepts of Epidemiology - An Integrated Introduction to the Ideas, Theories, Principles and Methods of Epidemiology. (3rd Edition). Oxford University Press, New York.

2.Celentano D. D. and Szklo M. (2018). Gordis Epidemiology. (6th Edition). Elseiver, USA.

3.Cheesbrough, M. (2004). District Laboratory Practice in Tropical Countries - Part 2, (2nd Edition). Cambridge University Press.

4.Ryan K. J. and Ray C. G. (2004). Sherris Medical Microbiology. (4th Edition), McGraw Hill, New York.

5.Topley W.W. C., Wilson, G. S., Parker M. T. and Collier L. H. (1998). Principles of Bacteriology. (9th Edition). Edward Arnold, London.

Web Resources

1. https://www.scielo.br/j/rbca/a/mjDFGTtfWtBm786ZmR9TG9d/?lang=en

2.https://hal.archives-ouvertes.fr/hal-00902711/document

3. https://www.who.int/csr/resources/publications/whocdscsreph200212.pdf

4.https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7187955/

5.https://www.who.int/diseasecontrol_emergencies/publications/idhe_2009_london_outbreaks.pdf

Mapping with Program Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M													
CO2				L	L	S								
CO3	M				S									
CO4					S									
CO5				S	S									

CLINICAL AND DIAGNOSTIC MICROBIOLOGY

Course Code: P2R3MBDSE3:2	**Credit:** 3
Category: Discipline Specific Elective	**Hrs/Week:** 4, **Total Inst.Hrs:** 60
Nature of the Course: Entrepreneurship	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Describe appropriate safety protocol and laboratory techniques for handling specimens and biomedical waste management.
2. Develop working knowledge of techniques used to identify infectious agents in the clinical microbiology lab.
3. Elucidate various diagnostic procedures in microbiology.
4. Acquire knowledge on different methods employed to check antibiotic sensitivity.
5. Gain knowledge on hospital acquired infections and their control measures.

Unit I: **12 Hrs**

Microbiology Laboratory Safety Practices -General Safety Guidelines, Handling of Biological Hazards, Infectious health care waste disposal - Biomedical waste management, Emerging and Re-emerging infections.

Unit II: **12 Hrs**

Diagnostic procedures - General concept of Clinical specimen collection, transport, storage and general processing in Microbiology laboratory - Specimen acceptance and rejection criteria.

Unit III: **12 Hrs**

Diagnosis of microbial diseases - Clinical, differential, Microbiological, immunological and molecular diagnosis of microbial diseases. Modern and novel microbial diagnostic methods. Automation in Microbial diagnosis.

Unit IV: **12 Hrs**

Antibiotic sensitivity tests - Disc diffusion - Stokes and Kirby Bauer methods, E test - Dilution - Agar dilution & broth dilution - MBC/MIC - Quality control for antibiotics and standard strains.

Unit V: **12 Hrs**

Nosocomial infections – common types, sources, reservoir and mode of transmission, pathogenesis and control measures. Hospital Infection Control Committee (HICC) – Functions.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Apply Laboratory safety procedures and hospital waste disposal strategies.	PO5, PO6, PO7
CO2	Collect various clinical specimens, handle, preserve and process safely.	PO6, PO7
CO3	Identify the causative agents of diseases by conventional and molecular methods following standard protocols.	PO6, PO7, PO9, PO11
CO4	Assess the antimicrobial susceptibility pattern of pathogens.	PO7, PO9
CO5	Trace the sources of nosocomial infection and recommend control measures.	PO5, PO7

Text Books

1.Collee J. G., Fraser A.G. Marmion B. P. and Simmons A. (1996). Mackie & McCartney Practical Medical Microbiology. (14th Edition). Elsevier, New Delhi. ISBN-10:0443047219 / ISBN-13-978-0443047213.

2.Tille P. M. (2021). Bailey and Scott's Diagnostic Microbiology. (15th Edition). Elsevier. ISBN:9780323681056.

3.Jawetz E., Melnick J. L. and Adelberg E. A. (2000). Review of Medical Microbiology. (19th Edition). Lange Medical Publications, U.S.A.

4.Mukherjee K.L. (2000). Medical Laboratory Technology.Vol. 1-3. (2nd Edition). Tata McGraw-Hill Education. ISBN-10:0074632604.

5.Sood R. (2009). Medical Laboratory Technology – Methods and Interpretations. (6th Edition). Jaypee Brothers Medical Publishers (P) Ltd. New Delhi. ISBN:9788184484496.

References Books

1.Murray P. R., Baron E. J., Jorgenson J. H., Pfaller M. A. and Yolken R.H. (2003). Manual of Clinical Microbiology. (8th Edition). American Society for Microbiology, Washington, DC. ISBN:1-555810255-4.

2.Bennett J. E., Dolin R. and Blaser M. J. (2019). Principles and Practice of Infectious Diseases. (9th Edition). Elsevier. EBook ISBN:9780323550277. Hardcover ISBN:9780323482554.

3.Ridgway G. L., Stokes E. J. and Wren M. W. D. (1987). Clinical Microbiology 7th Edition. Hodder Arnold Publication. ISBN-10:0340554231 / ISBN-13:9780340554234.

4.Koneman E.W., Allen S. D., Schreckenberg P. C. and Winn W. C. (2020). Koneman's Color Atlas and Textbook of Diagnostic Microbiology. (7th Edition). Jones & Bartlett Learning. ISBN:1284322378 9781284322378.

5.Cheesbrough, M. (2004). District Laboratory Practice in Tropical Countries - Part 2, (2nd Edition). Cambridge University Press. ISBN-13:978-0-521-67631-1 / ISBN-10:0-521-67631-2.

Web Resources

1.https://www.ncbi.nlm.nih.gov/books/NBK20370/

2.https://www.msdmanuals.com/en-in/home/infections/diagnosis-of-infectious3disease/diagnosis-of-infectious-disease

3.https://journals.asm.org/doi/10.1128/JCM.02592-20

4.https://www.sciencedirect.com/science/article/pii/S2221169116309509

5.http://www.textbookofbacteriology.net/normalflora_3.html

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1					S	M	M							
CO2						M	S							
CO3						M	S		M		S			
CO4							S		M					
CO5					S		M							

BIOREMEDIATION

Course Code: P2R3MBDSE3:3	**Credit:** 3
Category: Discipline Specific Elective	**Hrs/Week:** 4, **Total Inst.Hrs:** 60
Nature of the Course: Employability	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Describe the nature and importance of bioremediation and use in real world applications.
2. Describe the typical composition of waste water and application of efficient technologies for water treatment.
3. Explain the fundamentals of treatment technologies and the considerations for its design and implementation in treatment plants.
4. Explain the potential of microbes in ore extraction and acquaint students with methods of reducing health risks caused by xenobiotics.
5. Familiarize the role of plants and their associated microbes in remediation and management of environmental pollution.

Unit I: **12 Hrs**

Bioremediation - process and organisms involved. Bioaugmentation - Ex-situ and in-situ processes; Intrinsic and engineered bioremediation. Major pollutants and associated risks; organic pollutant degradation. Microbial aspects and metabolic aspects. Factors affecting the process. Recent developments and significance.

Unit II: **12 Hrs**

Microbes involved in aerobic and anaerobic processes in nature. Water treatment - BOD, COD, dissolved gases, removal of heavy metals, total organic carbon removal. Secondary waste water treatments - use of membrane bioreactor. Aquaculture effluent treatment. Aerobic sludge and landfill leachate process. Aerobic digestion.

Unit III: **12 Hrs**

Composting of solid wastes, anaerobic digestion - methane production and important factors

involved, Pros and cons of anaerobic process, sulphur, iron and nitrate reduction, hydrocarbon degradation, degradation of nitroaromatic compounds. Bioremediation of dyes, bioremediation in paper and pulp industries. Aerobic and anaerobic digesters – design. Various types of digester for bioremediation of industrial effluents.

Unit IV: **12 Hrs**

Microbial leaching of ores - process, microorganisms involved and metal recovery with special reference to copper and iron. Biotransformation of heavy metals and xenobiotics. Petroleum biodegradation - reductive and oxidative. Dechlorination. Biodegradable of plastics and super bug.

Unit V: **12 Hrs**

Phytoremediation of heavy metals in soil - Basic principles of phytoremediation - Uptake and transport, Accumulation and sequestration. Phytoextraction. Phytodegradation. Phytovolatilization. Rhizodegradation. Phytostabilization – Organic and synthetic amendments in multi metal contaminated mine sites. Role of Arbuscular mycorrhizal fungi and plant growth promoting rhizobacteria in phytoremediation.

Course Outcomes		
Course Outcomes		
CO1	Differentiate Ex-situ bioremediation and In-situ bioremediation. Assess the roles of organisms in bioremediation.	PO1, PO2, PO4, PO5
CO2	Distinguish microbial processes necessary for the design and optimization of biological processing unit operations.	PO1, PO4, PO5, PO11
CO3	Identify, formulate and design engineered solutions to environmental problems.	PO5, PO7, PO8, PO11
CO4	Explore microbes in degradation of toxic wastes and playing role on biological mechanisms.	PO5, PO6, PO7, PO8, PO9
CO5	Establish the mechanisms of Arbuscular mycorrhizal fungi and Plant growth promoting *Rhizobacteria* in phytoremediation.	PO1, PO5, PO6, PO7, PO8

Text Books

1.Bhatia H.S. (2018). A Text book on Environmental Pollution and Control. (2nd Edition). Galgotia Publications.

2.Chatterjee A. K. (2011). Introduction to Environmental Biotechnology. (3rd Edition). Printice-Hall, India.

3.Pichtel, J. (2014). Waste Management Practices: Municipal, Hazardous, and Industrial, 2nd edition, CRC Press.

4.Liu, D.H.F and Liptak, B.G (2005). Hazardous Wastes and Solid Wastes, Lewis Publishers..

5.Rajendran, P. & Gunasekaran, P. (2006). Microbial Bioremediation. 1st edition. MJP Publishers

References Books

1.Sangeetha J., Thangadurai D., David M. and Abdullah M.A. (2016). Environmental Biotechnology: Biodegradation, Bioremediation, and Bioconversion of Xenobiotics for Sustainable Development. (1st Edition). Apple Academic Press.

2.Singh A. and Ward O. P. (2004). Biodegradation and Bioremediation. Soil Biology. Springer.

3.Singh A., Kuhad R. C., and Ward O. P. (2009). Advances in Applied Bioremediation (1st Edition). Springer-Verlag Berlin Heidelberg, Germany.

4.Atlas, R.M & Bartha, R. (2000). Microbial Ecology. Addison Wesley Longman Inc.

5.Rathoure, A.K. (Ed.). (2017). Bioremediation: Current Research and Applications. 1st edition. I.K. International Publishing House Pvt. Ltd.

Web Resources

1.Bioremediation- Objective, Principle, Categories, Types, Methods, Applications (microbenotes.com)

2.https://agris.fao.org › agris-search

3.https://www.sciencedirect.com/topics/earth-and-planetary-sciences/bioremediation

4.https://www.intechopen.com/chapters/70661

5.https://microbiologysociety.org/blog/bioremediation-the-pollution-solution.html

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S	M		M	S									
CO2	S			M	S						S			
CO3					S		S	S			S			
CO4					S	S	S	S	S					
CO5	M				S	M	S	S						

BIOINFORMATICS

Course Code: P2R3MBDSE4:1	**Credit:** 3
Category: Discipline Specific Elective	**Hrs/Week:** 4, **Total Inst.Hrs:** 60
Nature of the Course: Employability	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Discuss about various biological data mining concepts, tools.
2. Elucidate the principles and applications of sequence alignment methods and tools.
3. Demonstrate different phylogenetic tree construction methods and its uses in phylogenetic analysis.
4. Acquaint with various approaches in predicting 3D and 2D structure of proteins.
5. Describe various tools and techniques used in molecular docking, immunoinformatics and subtractive genomics.

Unit I: **12 Hrs**

Biological Data Mining – Exploration of Data Mining Tools. Cluster Analysis Methods. Data Visualization. Biological Data Management. Biological Algorithms – Biological Primary and Derived Databases. Concept of Alignment, Pairwise Sequence Alignment (PSA), Multiple Sequence Alignment (MSA), BLAST, CLUSTALW, Scoring Matrices, Percent Accepted Mutation (PAM), Blocks of Amino Acid Substitution Matrix (BLOSUM).

Unit II: **12 Hrs**

Phylogenetic Tree Construction - Concept of Dendrograms. Evolutionary Trees - Distance Based Tree Reconstruction - Ultrametric trees and Ultrametric distances – Reconstructing Trees from Additive Matrices - Evolutionary Trees and Hierarchical Clustering - Character Based Tree Reconstruction - Maximum Parsimony Method, Maximum likelihood method - Reliability of Trees – Substitution matrices – Evolutionary models.

Unit III: **12 Hrs**

Computational Protein Structure prediction – Secondary structure – Homology modelling- Fold recognition and ab initio 3D structure prediction – Structure comparison and alignment –

Prediction of function from structure. Geometrical parameters – Potential energy surfaces – Hardware and Software requirements-Molecular graphics – Molecular file formats- Molecular visualization tools.

Unit IV: **12 Hrs**

Prediction of Properties of Ligand Compounds – 3D Autocorrelation -3D Morse Code-Conformation Dependent and Independent Chirality Codes –Comparative Molecular Field Analysis – 4 D QSAR –HYBOT Descriptors – Structure Descriptors – Applications – Linear Free Energy Relationships – Quantity Structure - Property Relationships –Prediction of the Toxicity of Compounds.

Unit V: **12 Hrs**

Molecular Docking- Flexible - Rigid docking- Target- Ligand preparation- Solvent accessibility- Surface volume calculation, Active site prediction- Docking algorithms- Genetic, Lamarckian - Docking analyses- Molecular interactions, bonded and nonbonded - Molecular Docking Software and Working Methods. Genome to drug discovery – Subtractive Genomics – Principles of Immunoinformatics and Vaccine Development.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Access to databases that provides information on nucleic acids and proteins.	PO1, PO4, PO6, PO7, PO9, PO10, PO13
CO2	Invent algorithms for sequence alignment.	PO7, PO9, PO10, PO13
CO3	Construct phylogenetic tree.	PO6, PO9, PO10
CO4	Predict the structure of proteins.	PO4, PO6, PO7, PO9, PO13
CO5	Design drugs by predicting drug ligand interactions and molecular docking.	PO4, PO5, PO6, PO7, PO9, PO10, PO13

Text Books

1.Lesk A. M. (2002). Introduction to Bioinformatics. (4th Edition). Oxford University Press.

2.Lengauer T. (2008). Bioinformatics- from Genomes to Therapies (Vol-1).Wiley- VCH.

3.Rastogi S. C., Mendiratta N. and Rastogi P. (2014). Bioinformatics - Methods and Applications (Genomics, Proteomics and Drug Discovery) (4th Edition). Prentice-Hall of India Pvt.Ltd.

4.Attwood, T.K. and Parry-Smith, D.J. (1999). Introduction to Bioinformatics. Addision Wesley Longman Limited, England.

5.Mount D.W., (2013).Bioinformatics sequence and genome analysis, 2ndedn.CBS Publishers, New Delhi.

References Books

1.Baxevanis A. D. and Ouellette F. (2004). Bioinformatics: A Practical Guide to the Analysis of Genes and Proteins. (2nd Edition). John Wiley and Sons.

2.Bosu O. and Kaur S. (2007). Bioinformatics - Database, Tools, and Algorithms. Oxford University Press.

3.David W. M. (2001). Bioinformatics Sequence and Genome Analysis (2nd Edition). CBS Publishers and Distributors(Pvt.)Ltd.

4.Xiong J, (2011). Essential bioinformatics, First south Indian Edition, Cambridge University Press.

5.Harshawardhan P.Bal, (2006). Bioinformatics Principles and Applications, Tata McGraw-Hill Publishing Company Limited.

Web Resources

1.https://www.hsls.pitt.edu/obrc/

2.https://www.hsls.pitt.edu/obrc/index.php?page=dna

3.https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1669712/

4.https://www.ebi.ac.uk/

5.https://www.kegg.jp/kegg/kegg2.html

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M			M		M			M	M			M	
CO2							S		S	S			S	
CO3						S			S	S				
CO4				S		S	S		S				S	
CO5				S	S	S	S		S	S			S	

NANOBIOTECHNOLOGY

Course Code: P2R3MBDSE4:2	**Credit:** 3
Category: Discipline Specific Elective	**Hrs/Week:** 4, **Total Inst.Hrs:** 60
Nature of the Course: Skill Development	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Analyze nanomaterials based on the understanding of nanobiotechnology.
2. Discuss the methods of fabrication of nanomaterials.
3. Gain Knowledge on characterization of nanomaterials.
4. Discover nanomaterials for targeted drug delivery.
5. Explain nanomaterials in nanomedicine and environmental pollution.

Unit I: **12 Hrs**

Introduction to nanobiotechnology, Nano size-changing phenomena at nano scale, Classification of nanomaterials based on their dimensions (0D, 1D, 2D and 3D materials) and based on realization of their applications (The First, second, third and fourth generation materials),Class of nanomaterials and their applications. Need for nanomaterials and the risks associated with the materials.

Unit II: **12 Hrs**

Fabrication of Nanomaterials-Top-down and Bottom-up approaches, Solid phase synthesis-milling, Liquid phase synthesis-Sol-gel synthesis, colloidal synthesis, micro emulsion method, hydrothermal synthesis and solvo thermal synthesis, Vapour/Gas phase synthesis-Inert gas condensation, flame pyrolysis, Laser ablation and plasma synthesis techniques. Microbial synthesis of nanoparticles.

Unit III: **12 Hrs**

Characterization of nanoparticles – Based on particle size/morphology- Dynamic light scattering (DLS),Scanning electron microscopy (SEM), Transmission electron microscopy (TEM), Atomic force microscopy(AFM), Based on surface charge-zeta potential, Based on structure –X-ray diffraction (XRD), Fourier transform infrared spectroscopy (FTIR), Energy dispersive X-ray

analysis (EDX),Based on optical properties- UV – Spectrophotometer, Based on magnetic properties-Vibrating sample magnetometer(VSM).

Unit IV: **12 Hrs**

Nanomaterial based Drug delivery and therapeutics-surface modified nano particles, MEMS/NEMS based devices, peptide/DNA coupled nanoparticles, lipid and inorganic nano particles for drug delivery, Metal/metaloxide nano particles as antibacterial, antifungal and antiviral agents. Toxicity of nanoparticles and Toxicity Evaluation.

Unit V: **12 Hrs**

Nanomaterials in diagnosis-Imaging, nanosensors in detection of pathogens. Treatment of surface water, ground water and waste water contaminated by toxic metal ions, organic and inorganic solutes and microorganisms.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Employ knowledge in the field of nanobiotechnology for development.	PO1, PO9
CO2	Identify various applications of nanomaterials in the field of medicine and environment.	PO1, PO9
CO3	Examine the prospects and significance of nanobiotechnology.	PO1, PO6, PO11
CO4	Identify recent advances in this area and create a career or pursue research in the field.	PO1, PO5, PO7, PO9
CO5	Design non-toxic nanoparticles for targeted drug delivery.	PO1,PO5, PO7, PO9, PO11

Text Books

1.Brydson R. M., Hammond, C. (2005). Generic Methodologies for Nanotechnology: Characterization. In Nanoscale Science and Technology. John Wiley & Sons, Ltd.

2.Leggett G. J., Jones R. A. L. (2005). Bionanotechnology. In Nanoscale Science and Technology. John Wiley & Sons, Ltd.

3. Mohan Kumar G. (2016). Nanotechnology: Nanomaterials and nanodevices. Narosa Publishing House.

4.Goodsell D. S. (2004). Bionanotechnology. John Wiley & Sons, Inc.

5.Pradeep T. (2007). Nano: The Essentials-Understanding nanoscience and nanotechnology. Tata McGraw-Hill.

References Books

1.Nouailhat A. (2008). An Introduction to Nanoscience and Nanotechnology, Wiley.

2.Sharon M. and Maheshwar (2012). Bio-Nanotechnology: Concepts and Applications. New Delhi. Ane books Pvt Ltd.

3. Niemeyer C.M. and Mirkin C. A. (2005). Nanobiotechnology. Wiley Interscience.

4.Rehm, B. (2006). Microbial Bionanotechnology: Biological Self-Assembly Systems and Biopolymer-Based Nanostructures. Horizon Scientific Press.

5.Reisner, D.E. (2009). Bionanotechnology: Global Prospects. CRC Press

Web Resources

1.https://www.gale.com/nanotechnology

2.https://www.understandingnano.com/resources.html

3.http://dbtnanobiotech.com/index2.php

4.http://www.istl.org/11-winter/internet1.html

5.https://www.cdc.gov/niosh/topics/nanotech/default.html

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S			M					M					
CO2	S								S					
CO3	S					M					S			
CO4	S				S		M		S					
CO5	S				S		M		S		S			

CLINICAL RESEARCH AND CLINICAL TRIALS

Course Code: P2R3MBDSE4:3	**Credit:** 3
Category: Discipline Specific Elective	**Hrs/Week:** 4, **Total Inst.Hrs:** 60
Nature of the Course: Employability	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Provide an overview of history and methods involved in conducting clinical research.
2. Design the principles involved in ethical, legal, and regulatory issues in clinical research on human subjects.
3. Describe principles and issues involved in monitoring patient-oriented research.
4. Formulate a well- defined quality assurance and quality control plans.
5. Acquire business development skills in the area of clinical research.

Unit I: **12 Hrs**

Introduction to Clinical Research: Clinical Research: An Overview, Different types of Clinical Research. Clinical Pharmacology: Pharmacokinetics, Pharmacodynamics, Pharmacoepidemiology, Bioavailability, Bioequivalence, Terminologies and definition in Clinical Research. Drug Development Process: Drug Discovery Pipeline, Drug Discovery Process. Preclinical trail, Human Pharmacology (Phase-I), Therapeutic Exploratory trail (Phase-II), Therapeutic Confirmatory Trail (Phase-III) and Post marketing surveillance (Phase-IV).

Unit II: **12 Hrs**

Ethical Considerations and Guideline in Clinical Research: Historical guidelines in Clinical Research-Nuremberg code, Declaration of Helsinki, Belmont report. International Conference on Harmonization (ICH)-Brief history of ICH, Structure of ICH & ICH Harmonization Process, Guidelines for Good Clinical Practice. Regulation in Clinical Research-Drug and cosmetic act, FDA, Schedule-Y- Ethics Committee and their responsibilities. Clinical Research Regulatory Submission & approval Process- IND, NDA and ANDA submission Procedure. DCGI submission procedure. Other Regulatory authorities- EMEA, MHRA, PhRMA.

Unit III: **12 Hrs**

Clinical Trial Management: Key Stakeholders in Clinical Research, Ethics Committees and Institutional Review Board, Responsibilities of Sponsor. Responsibilities of Investigator, Protocol in Clinical Research Clinical Trial Design, Project Planning Project Managements - Informed Consent, Investigator's Brochure (IB), Selection of an Investigator and Site, Patient screening, Inclusion and exclusion criteria, Randomization, Blinding. Essential Documents in clinical research -IB, ICF, PIS, TMF, ISF, CDA & CTA.

Unit IV: **12 Hrs**

Quality Assurance, Quality Control & Clinical Monitoring: Defining the terminology-Quality, Quality system, Quality Assurance & Quality Control-QA audit plan. 21 CRF Part 11, Site Auditing, Sponsor Compliance and Auditing, SOP For Clinical Research-CRF Review & Source Data Verification, Drug Safety Reporting Corrective and preventative action process.

Unit V: **12 Hrs**

Business Development in the Clinical Research Industry: Introduction & Stages of Business Development-Start-up Phase, Growth Phase, Maturity Phase, Decline Phase. Outsourcing in Clinical Research, Reasons for outsourcing to contract research organizations, The India Advantage, Scope and Future of CRO, List of Clinical Research Organizations in India, List of IT companies offering services in Clinical Research. Role of business development manager.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Apprehend the Drug Development process and different phases of clinical trials.	PO1, PO2, PO3, PO5
CO2	Recognize the ethics and regulatory perspectives on clinical research trials activities.	PO3, PO5, PO6, PO9
CO3	Accentuate about clinical trials management concepts and documentation process.	PO2, PO4, PO6, PO9
CO4	Accomplish quality assurance and quality control to ensure the protection of human subjects and the reliability of clinical trial results.	PO2, PO4. PO6. PO7, PO9
CO5	To nurture skills recitation to commercial start up and industriousness.	PO4, PO8, PO9, PO11, PO13

Text Books

1.Gallin J. I., Ognibene F. P. and Johnson L. L. (2007). Principles and Practice of Clinical Research. (4th Edition). Elsevier, 2007.ISBN-10: 0128499052

2.Friedman L. M., Furberg C. D. and Demets D. (1998). Fundamentals of Clinical Trials, Vol: XVIII. (3rd Edition). Springer Science & Business Media.

3.Hulley S. B., Cummings S. R., Browner W. S., Grady D. G. and Newman T. B. (2013). Designing Clinical Research. (4th Edition). Jaypee Medical. ISBN-13: 978-1608318049.

4.Reed,G. (2004). Prescott and Dunn's Industrial Microbiology, 4th edn, CBS publication and distributors.

5.Himanshu B. Text book of Clinical Research, Pee Vee books.

References Books

1.Friedman L.M., Fuberge C.D., DeMets D. and Reboussen, D.M. (2015). Fundamentals of Clinical Trials, Springer.

2.Browner W. S., (2012). Publishing and Presenting Clinical Research. (3rd Edition). Lippincott Williams and Wilkins.

3.Rondel R. K., Varley S. A. and Webb C. F. (2008). Clinical Data Management. (2nd Edition). Wiley.

4.Peppler, H.J. and Pearl Man, D. (1979). Fermentation Technology, Vol 1 & 2, 2nd Edition Academic Press, London.

5.E1-Mansi, E.M.T., Bryce, C.F.A., Demain, A.L. and Allman,A.R. (2007). Fermentation Microbiology and Biotechnology. 2nd Edition, CRC press, Taylor and Francis Group.

Web Resources

1. https://www.hzu.edu.in/uploads/2020/10/Textbook-of-Clinical-Trials-Wiley-(2004).pdf

2.https://www.routledge.com/A-Practical-Guide-to-Managing-Clinical-Trials/Pfeiffer-Wells/p/book/9780367497828

3.https://www.auctoresonline.org/journals/clinical-research-and-clinical-trials

4.https://www.who.int/health-topics/clinical-trials#tab=tab_1

5.https://www.cancerresearchuk.org/about-cancer/find-a-clinical-trial/what-clinical-trials-are/types-of-clinical-trials

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S	S	S		S									
CO2			S		S	S			S					
CO3		S		S		S			S					
CO4		S		S		S	S		S					
CO5				S				S	S		S		M	

VERMITECHNOLOGY

Course Code: P2R3MBSEC1:1	**Credit:** 2
Category: Skill Enhancement Course 1	**Hrs/Week:** 4, **Total Inst.Hrs:** 60
Nature of the Course: Entrepreneurship	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. Introduce the concepts of vermicomposting.
2. Explain the physiology, anatomy and biology of earthworms.
3. Acquire the knowledge of the vermicomposting process.
4. Explain the trouble shooting, harvesting and packaging of vermin composts.
5. Gain knowledge on applications of vermin composts and their value added products.

Unit I: 12 **hrs**

Introduction to Vermiculture - Definition, classification, history, economic importance- In sustainable agriculture, organic farming, earthworm activities, soil fertility & texture, soil aeration, water impercolation, decomposition & moisture, bait & food and their value in maintenance of soil structure. Its role in the bio transformation of the residues generated by human activity and production of organic fertilizers. Choosing the right worm. Useful species of earthworms. Local species of earthworms. Exotic species of earthworms. Factors affecting distribution of earthworms in soil.

Unit II: 12 **hrs**

Earthworm Biology and Rearing - Key to identify the species of earthworms. Biology of *Eisenia fetida.* a) Taxonomy Anatomy, physiology and reproduction of Lumbricidae. b) Vital cycle of *Eisenia fetida*: alimentation, fecundity, annual reproducer potential and limiting factors (gases, diet, humidity, temperature, PH, light, and climatic factors). Biology of *Eudrilus eugeniae*. c) Taxonomy Anatomy, physiology and reproduction of Eudrilidae. d) Vital cycle of *Eudrilus eugeniae*: alimentation, fecundity, annual reproducer potential and limit factors (gases, diet, humidity, temperature, PH, light, and climatic factors).

Unit III: 12 **hrs**

Vermicomposting Process - Feeds for Vermitech systems- Animal manures- Kitchen Waste and Urban waste- Paper pulp and card board solids- Compost and waste products- Industrial Wastes. Vermicomposting Basic process- Initial pre-composting phase- Mesophilic phase- Maturing and stabilization phase- Mechanism of Earthworm action. Methods of vermicomposting- a) windrows system; b) wedge system; c) container system-pits, tanks & cement rings; commercial model; beds or bins-top fed type, stacked type, d) Continuous flow system.

Unit IV: 12 **hrs**

Vermicomposting - Trouble Shooting-Temperature-Aeration- Acidity- Pests and Diseases- Ants, rodents, Birds, Centipedes, sour crop, Mite pests. Odour problems. Separation techniques- Light Separation-Sideways Separation-Vertical Separation-Gradual transfer. Harvesting Earthworms- manual method- migration method. Packing & Nutritional analysis of vermicompost.

Unit V: 12 **hrs**

Applications of Vermiculture - Vermiculture Bio-technology, use of vermi castings in organic farming/horticulture, as feed/bait for capture/culture fisheries; forest regeneration. Application quantity of vermicompost in Agricultural fields- crops, fruits, vegetables & flowers. By-products and value-added products- Verm wash- vermicompost tea-vermi meal-enriched vermicompost- pelleted vermicompost.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Compare and contrast the uses of vermicompost to the soil.	PO1, PO4, PO5, PO9,
CO2	Recommend different species of earthworms after acquiring knowledge on its biology.	PO1, PO4, PO6, PO9
CO3	Design the vermicomposting process.	PO1, PO4, PO6, PO7, PO8
CO4	Assess the Best Practices of Vermicomposting	PO6,PO7, PO8,PO9,
CO5	Recommend the applications of vermicompost to different soils and for different crops.	PO1, PO4, PO5,PO6, PO7

Text Books

1. Ismail S. A. (2005). The Earthworm Book, Second Revised Edition. Other India Press, Goa, India.
2. Rathoure A. K., Bharati P. K. and Ray J. (2020). Vermitechnology, Farm and Fertilizer. Vermitechnology, Farm and Fertilizer Discovery Publishing House Pvt Ltd.
3. Christy M. V. 2008. Vermitechnology, (1st Edition), MJP Publishers.
4. The complete technology book on Vermiculture and Vermicompost with manufacturing Process, machinery equipment details and Plant Layout. AB Press.
5. Keshav Singh (2014). A Textbook of vermicompost: Vermiwash and Biopesticide.

Reference Books

1. Roy D. (2018). Handbook of Vermitechnology. Lambert Academic Publishing.
2. Kumar A. (2005). Verms and Vermitechnology, A.P.H. Publishing Corporation, New Delhi.
3. Lekshmy M. S., Santhi R. (2012). Vermitechnology, Sara Publications, New Delhi, India. Edwards CA, Arancon NQ ShermanRL. (2011) Vermiculture Technology: Earthworms, Organic Wastes, and Environmental Management 1st edn.CRC Press.
4. Ismail, S.A. (1997). Vermicology-The Biology of Earthworm.1st edn. Orient longman.

Web Resources

1. https://en.wikipedia.org/wiki/Vermicompost

   http://stjosephs.edu.in/upload/papers/9567411a78c63d4ccfbbe85e6aa22840.pdf
2. https://www.kngac.ac.in/elearning-portal/ec/admin/contents/4_18K4ZEL02_2021012803204629.pdf
3. https://composting.ces.ncsu.edu/vermicomposting-2/
4. https://rodaleinstitute.org/science/articles/vermicomposting-for-beginners/

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S			M	S				S					
CO2	S			M		S			S					
CO3	S			S		S	S	S						
CO4						S	S	S	S					
CO5	S			M	S	M	S							

MUSHROOM TECHNOLOGY

Course Code: P2R3MBSEC1:2	**Credit:** 2
Category: Skill Enhancement Course	**Hrs/Week:** 4, **Total Inst.Hrs:** 60
Nature of the Course: Entrepreneurship	**Marks:** CIA: 25+ EXT: 75 = 100

Course Objectives

1. To give the basics of mushrooms in human food
2. To know mushroom propagation for food industries
3. To give an opportunity for future entrepreneurship
4. To get knowledge on mushroom cultivation
5. To get idea on nutrition availability in mushroom

Unit I: 12 **hrs**

Introduction – History – scope of edible mushroom cultivation – Types of edible mushroom available in India – *Calacybe indica, Volvariella volvacea, Pleurotussajor-caju. Agaricus bisporus.*

Unit II: 12 **hrs**

Pure culture – preparation of media (PDA and Oatmeal agar media) sterilization – Preparation of test tube slants to store mother culture – culturing of Pleurotus mycelium on petriplates – Preparation of mother spawn in saline bottle and polypropylene bags and their multiplication.

Unit III: 12 **hrs**

Cultivation Technology: Infra structure, locally available substrates, polythene bags, vessels, inoculation hood, inoculation loop, low cost stove, sieves, Culture rack, Mushroom unit (Thatched house) – Mushroom bed preparation – Paddy straw, sugarcane trash, maize straw, banana leaves.

Unit IV: 12 **hrs**

Storage and nutrition: Short term storage – long term storage (scanning, pickles, papads, drying, and storage in salt solutions) – Nutrition: Proteins, amino acids, mineral elements. Nutrition: Carbohydrates – Crude fiber content, vitamins.

Unit V: 12 **hrs**

Food preparation: Types of foods prepared from mushroom – soup, cutlet, omlette, samosa, pickles, curry. Research centers – National level and Regional level. Cost benefit ratio – Marketing in India and abroad – Export value.

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Gain Knowledge on mushroom cultivation techniques so that they can become employable in agro-based industries.	PO1, PO4, PO5, PO9,
CO2	Basic idea on training to go for self-employment	PO1, PO4, PO6, PO9
CO3	Seek information on difference between edible mushroom and poisonous mushroom	PO1, PO4, PO6, PO7, PO8
CO4	Get educated on food preparation using mushroom	PO6,PO7, PO8,PO9,
CO5	Obtain knowledge on nutrition in mushroom	PO1, PO4, PO5,PO6, PO7

Text Books

1. Nita Bahl (1988) Hand Book of Mushrooms, II edition Vol I & II.
2. Shu Ting Chang, Philip G. Miles, Chang. S.T. (2004). Mushrooms: Cultivation, nutritional value, medicinal effect and environmental impact. 2nd edition, CRC press.
3. Paul Stamets, J.S. and Chittom, J.S. (2004). Mushroom Cultivator: A Practical guide to grow mushrooms at home, Agarikon press.

Reference Books

1. Marimuthu*et al.*, (1991) Oyster Mushrooms, Dept. of Plant Pathology, TNAU, Coimbathore.
2. Tewari and Pankaj Kapoor S.C. (1988) Mushroom cultivation, Mittal Publications, Delhi.

3. Swaminathan M. (1990) Food and Nutrition. Bappco. The Bangalore Pritning and Publishing Company Ltd., Bangalore.

Web Resources

1. https://www.sciencedirect.com/science/article/pii/B9780120403073500081
2. https://justagriculture.in/files/newsletter/2021/june/02.%20MUSHROOM%20CULTIVATION%20TECHNOLOGY.pdf
3. https://www.drdo.gov.in/mushroom-cultivation-technology

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S			M	S				S					
CO2	S			M		S			S					
CO3	S			S		S	S	S						
CO4						S	S	S	S					
CO5	S			M	S	M	S							